

An ecophysiological model of *Alexandrium minutum*. Effect of phosphorus on vegetative growth.

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Introduction

Alexandrium spp are poison-producing dinoflagellate species known to produce near-shore blooms and harmful to shellfish production in harvesting bans. Along French coasts, *Alexandrium minutum* (Halim) blooms were first recorded in estuaries of northern Brittany in June 1988 and since, the phenomenon is believed to have spread. In the Penzé estuary, a tidal flat estuary connected to the west part of Morlaix bay (Northern Brittany, France), proliferation events are strongly linked to neap tides of late spring/early summer. There, cell density may reach high levels ($44.6 \cdot 10^6 \text{ cell.l}^{-1}$ up-estuary in June, 1997).

Two major biological processes are involved in these blooms : firstly, the synchronous germination of cysts from sediment beds ; secondly, the ability of *A. minutum* to compete for nutrients in nutrient enriched coastal waters. The Penzé is a river strongly eutrophicated where nitrogen is always in excess but phosphorus may become limiting. For a better understanding of *Alexandrium* blooms, a numerical model, based on experimental culture datas (Probert, 1999 ; Erard et al., 2003; Labry et al., 2004), is built in order to reproduce the vegetative growth linked to phosphorus disponibility.

Methods

The data set / The simulation proceeding

The Labry culture experiments were conducted on *A. minutum* (AM89BM) in filtered seawater under optimal environmental conditions ($T=18^\circ\text{C}$; $I=200 \mu\text{E.m}^{-2}.\text{s}^{-1}$; $f/2/4 \text{ NO}_3$) and no PO_4 .

Processes parametrization has been performed on semicontinuous cultures with a range from 0,05 to 0,5 dilution rate and a renewal medium containing $f/2/4 \text{ PO}_4$ ($9 \mu\text{M}$) and $f/2 \text{ NO}_3$ ($880 \mu\text{M}$). Model calibration has been performed on batch experiments with PO_4 enrichment with a $4 \mu\text{M}$ pulse at 1, 2, 3, 5, 7 and 10 days. Model validation has been performed on semicontinuous cultures with a 0,15 dilution rate for all parameters except PO_4 added each 1, 2, 4 or 6 days (in a corresponding input of $9 \mu\text{M}$ each day) and on batch culture from Erard (N, P concentrations related to *in situ* observations) and Probert experiments.

Processes has been adjusted using the Matlab interface (curvefit in Curve Fitting toolbox).

The numerical model has been written under Stella software.

Results

1- Growth and P uptake parametrization

Growth rate (μ) is derived from Droop (Droop, 1969) :

$$\mu = \mu_{\text{max}} * (1 - Q_{\text{pmin}}/Q_{\text{p}})$$

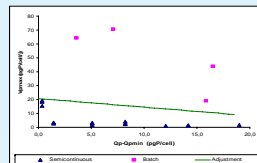


Figure 1 : Vmax parametrization

Uptake is adjusted following Labry experiments relating the P cell quota and Michaelis-Menten for $[\text{PO}_4]$ in the medium: $VP = VP_{\text{max}} * (P/(P+K_p))$. With $VP_{\text{max}} = -0,6183 * (Q_{\text{p}} - Q_{\text{pmin}}) + 20,67$ ($R^2=0,25$).

2- Model calibration on batch culture experiments

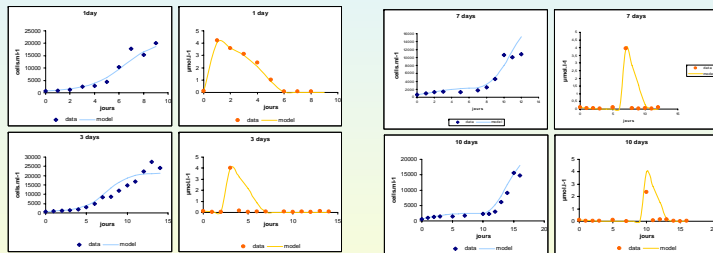


Figure 2 : Model Calibration on the Labry batch cultures. (*Alexandrium* in blue, PO_4 in red)

Parameters	Unit	Value
μ_{max}	Maximal growth rate	d^{-1} 1,22
Q_{pmin}	P minimum quota	pgP.cell^{-1} 6,49
V_{pmax}	P maximum uptake	$\text{pgP.cell}^{-1}.\text{d}^{-1}$ 20,67
KP	Half saturation constant for PO_4 uptake	mmol.m^{-3} 0,933

Table 1 : Parameters used in the model

2- Model validation on culture experiments

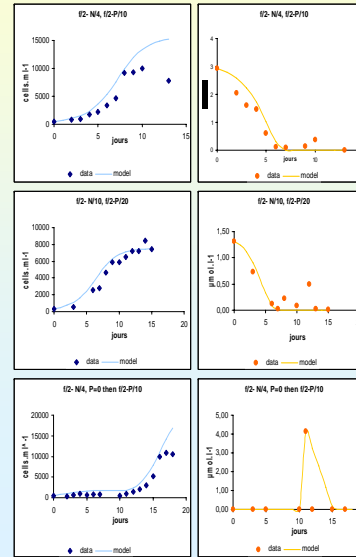


Figure 3 : Model Validation of Erard batch cultures. (*Alexandrium* in blue, PO_4 in red)
 $f/2-N/4$, $f/2-P/10$: *in situ* conditions observed in May 1997
 $f/-N/10$, $f/2-P/20$: minimal conditions observed *in situ*

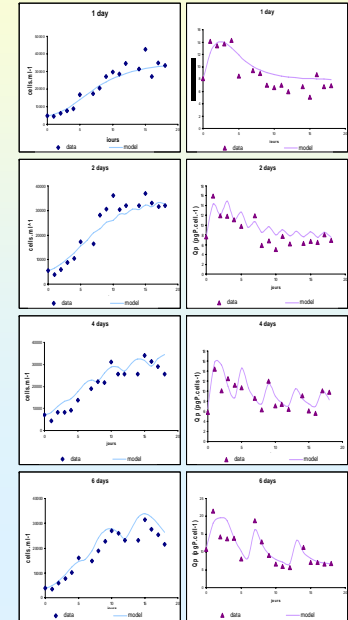


Figure 4 : Model Validation of Labry semicontinuous cultures. (*Alexandrium* in blue, P quota in violin)

Discussion

This model reproduces correctly both the population growth and the nutrient uptake in condition of P limitation for a large data set.

As it has been shown in the P uptake ajustement, it is difficult to reproduce the shift in P uptake for P limitation (low quota). This may be explained by the different evolution of these experiments. The ajustement may be not so adequate and it may explain, in the calibration as well as in the validation steps, the discrepancies between PO_4 simulated and data after PO_4 input in a limiting culture.

Validation on Erard cultures shows that the model surestimates the population yield. This may be due to an overestimate growth rate in this case as Erard found for these experiments a lower μ_{max} ($\mu_{\text{max}}=0,92 \text{ d}^{-1}$).

So, it may be important for future *Alexandrium* simulations (culture experiments or *in situ* simulations) to apply this model with a range for the parameters between parameters measurements and calibration.

Unit	Range
μ_{max}	d^{-1} 0,92 (Erard et al., 2003) - 1,44 (Probert, 1999)
Q_{pmin}	pgP.cell^{-1} 5 (Labry, compers.) - 17 (Probert, 1999)
V_{pmax}	$\text{pgP.cell}^{-1}.\text{d}^{-1}$ 20,4 - 71,4 (Labry et al. 2004)
KP	mmol.m^{-3} 0,28 - 1,62 (Labry et al., 2004)

Conclusion

This ecophysiological model of *Alexandrium minutum* takes into account the influence of PO_4 stress on vegetative growth. It has to be developed to take into account as well, nitrate, light and temperature control on vegetative growth as well as parametrization of sexual reproduction (see Chapelle et al., 2004). To understand *in situ* the occurrence of *Alexandrium* blooms, simulations should be performed coupled to an hydrodynamical model giving currents, water stability, and an ecological label describing nutrients and coexistence/competition with other planktonic algae.

References

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